Skin Lesion Surface Area and Erythema Intensity Calculation for PASI Determination using Stereo Vision

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Abstract—There are several tools to determine the severity of psoriasis and to track the effect of treatment, one of which is Psoriasis Area and Severity Index (PASI). PASI score calculation needs an accurate lesion area to total skin area ratio. To accurately determine these areas, 3D reconstruction from stereo vision has been used. Disparity maps and consecutively point clouds of the object of interest were made from stereo pictures. The object for testing this method has been the arm. Transforming the colour space from red, green and blue (RGB) to YCbCr (Y:luminance; Cb: chrominance-blue; Cr: chrominance-red) and inspecting the point clouds of each separate component, provides detection of red and lighter lesions. A point cloud of these lesions can then be made. Using Delaunay triangulation on the point clouds results in a mesh of which the area can be calculated. Tests have been conducted on light and dark skin shades, with red and pink lesions. It has also been tested for multiple lesions placed a few centimetres apart. The test results showed that the formulated method works on both light and dark shade skin for red lesions, and detection of multiple lesions can be done as well. The method does not perform as well on lighter, pink lesions. The calculated lesion areas differed to the actual areas with an average factor of +37.45%.

I. INTRODUCTION

Surface reconstruction is widely used for applications such as environment mapping, 3D modelling, and medical imaging. In medical applications, surface reconstruction is used to inspect and calculate the surface area of the skin, providing knowledge when treating certain diseases. For psoriasis which is mainly recognisable by red patches on the skin knowing the skin area is an important aspect for determining the Psoriasis Area and Severity Index (PASI) score, which indicates the severity of the lesions and serves as an indication on how well the treatment works [1]. Determining this area is still a tough task, and doing so by hand takes a lot of time. An existing method which computes this score, uses a mono camera [2]. It enables doctors to quickly obtain the surface area and also determines the severity of the other criteria needed to calculate the PASI score. Other existing methods for surface reconstruction use mono cameras as well, or methods such as photogrammetry, laser scanning, or millimeter wave scanning. With these methods, the surface area of the skin can be calculated to diagnose, monitor, or to keep track of treatments [3]. The existing surface reconstruction methods which do not use pictures, do not compute a full coloured mesh. This makes detecting different coloured areas - such as psoriasis lesions - challenging. Furthermore, using mono cameras does not provide area calculation in three dimensions, possibly

making the results less accurate than they could be. Existing methods also tend to be quite expensive, e.g. the reconstruction system at Robotics and Mechatronics (RaM) which costs over \in 70,000. Large camera setups could also make patients uncomfortable, which should be prevented if possible. This paper presents an affordable and handheld solution to surface calculation using stereo vision. As application, the method has been developed to detect lesions on different skin colours and calculate their area, making it useful for determining PASI scores. The lesion detection also determines the intensity of the redness.

II. MATERIALS

The following materials have been used for implementation and testing of this method:

- Stereo Camera: Non Distortion Dual Lens USB3.0 Camera Module Synchronization HD 960P OTG UVC Plug Play Driverless 3D VR Stereo Webcam (€75). Outputs stereo images with a resolution of 960x2560, i.e. 960x1280 for each separate left and right image.
- Software: MATLAB R2019a, including the Image Processing Toolbox.
- Lesion samples: red and pink stickers have been made, with areas of 12 cm² and 6 cm². The accuracy of the surface calculation will be determined by comparing this area to the calculated area.

III. METHODS

The PASI score is calculated by looking at four criteria per body part (head, arms, trunk and legs) [1], [2]:

- Area: the percentage of skin covered by the lesions, categorised as (0): 0%, (1): 1 9%, (2): 10 29%, (3): 30 49%, (4): 50 69%, (5): 70 89%, (6): 90 100%.
- Erythema: the redness of the lesions.
- Induration: the thickness of the lesions.
- Desquamation: the scaling of the lesions.

The bottom three criteria are categorised by five intensity scores: (0): Absent, (1): Mild, (2): Moderate, (3): Severe, (4): Very severe. This research will only present a solution to the first two criteria, the area and erythema of the lesion. These criteria can be determined by looking at the 3D model made from the stereo images. To make this 3D model, the process as seen in Figure 1 has been followed.



Fig. 1: Functional block diagram of the general process.

A. Make stereo pictures

Before being able to process stereo pictures, the camera has to be calibrated. This has been done with the Stereo Camera Calibrator app from MATLAB. After this, stereo pictures were made of an arm on which a lesion can be seen. For the tests, the lesion samples as described in Section II were examined.

B. Compute disparity map

From the taken stereo pictures, disparity maps were made by using MATLAB's Semi-Global Matching (SGM) function. SGM works especially well for fine structured objects and their borders and still works well under radiometric changes [4], [5]. It also provides less occlusions than Block Matching (BM), MATLAB's other disparity function. Disparity maps show the difference in distance between the left and right images; objects closer to the camera will have larger disparity than objects far away. Before being able to compute the disparity map, the disparity range should be determined. This has been done by drawing a line on the anaglyph image of each stereo pair, after which the range was automatically calculated. Doing so provides an accurate disparity range, such that it is correct for every stereo pair.

C. Obtain point cloud

Obtaining a correct, denoised point cloud takes a few steps. The disparity map should be correct, and not contain too many occlusions or noise, as the same noise present in the disparity map will be present in the point cloud as well. Removing the noise has been done by first down sampling the point cloud, then by looking at the distance between neighbouring points. If this distance is deemed too large, a point is considered noise. Another method is looking at the clusters, which are groups of points gathered in a certain area. Assuming the cluster of the arm is the largest one, smaller clusters are left out, leaving a denoised point cloud.

D. Mesh point cloud

For meshing the obtained volume point cloud, Delaunay triangulation has been used. The convex hull of the point cloud, or rather the mesh, in the x-y plane, consists of triangles formed by Delaunay edges [6]. Then the z coordinates of each point can be added, making the mesh three dimensional. However, using this method results in excess area, which should be left out when calculating the area of the arm and the lesion.

E. Calculate surface area

Since the mesh only consists of triangles, calculating the surface areas has been done by summing the area of the separate triangles. However, the triangulation method used here also makes triangles between outer points which are on opposite sides, since it is a convex hull. To solve this issue, the excess areas were detected by looking at their area and maximum edge length. These areas were not part of the summation.

F. Detect lesion

Using the YCbCr (Y: luminance; Cb: chrominance-blue; Cr: chrominance-red [7]) colour space deemed more reliable than the red, green and blue (RGB) colour space for detecting the lesions. By looking at an image in only the Cr component, the red areas - which have the highest Cr value - are highlighted. For lighter lesions the other two colours (Y and Cb) have been inspected as well. Since the area of the lesion should be determined from a 3D mesh, the lesion has been detected from the point cloud in the colour of one of the YCbCr components. A point cloud of only the lesion is left, which can be meshed and examined for area.

G. Determine erythema severity (redness)

The erythema severity of the lesion has been determined by comparing the redness of the stereo picture to the redness of the pictures as seen in Figure 2. To determine the redness, the Y, Cb, or Cr values are looked at, corresponding to which colour was used to detect the lesion. The colour values of the lesions were compared to those of the reference images. The severity is then the number of the picture the lesion corresponds to the most.



Fig. 2: Erythema intensity reference pictures [1].

H. Testing

To test the method, tests have been conducted on different skin colours, using the red and pink stickers as mentioned in Section II. Additionally, the method has been tested for two separated lesions. The calculated areas were compared to the actual measured areas of the stickers by calculating the difference with Equation 1. For each test, four images were taken from a distance of approximately 1 meter.

$$\frac{|area_{\rm real} - area_{\rm calculated}|}{area_{\rm real}} \cdot 100\%.$$
(1)

The PASI score has been determined for every test, using Equation 2. Assuming the sticker lesion is the only lesion on the whole body, the components regarding the head, trunk and legs are equal to zero and can be left out of the original equation [1].



Fig. 3: Results of test with red lesion on light skin tone, stereo pair 2.

$$PASI = (arms_{\text{erythema}} \cdot 0.2) \cdot arms_{\text{area}}, \tag{2}$$

where $arms_{erythema}$ and $arms_{area}$ are the intensity scores mentioned at the beginning of this section. To calculate the lesion to arm area ratio, the calculated arm area has been multiplied by four, to represent both arms from both front and back.

IV. RESULTS

The meshes presented in this section have red triangles in them. These triangles were areas not taken into account when calculating the total surface area. They have been automatically detected. The axes in the 3D models are defined as follows: *x*: the length, *y*: the width, *z*: the height/depth of the arm/lesion. Note that the axes are in decimeters.

For the first test with one red lesion on light skin, the end results as well as the intermediate results of stereo pair 2 can be seen in Figure 3. For the other tests, per skin tone and lesion colour, only the stereo image and surface mesh of the lesions of the test with the best result (the highlighted tests in Table I) are shown. The calculated areas and the difference to the actual area of all tests can be seen in Table I. The calculated areas of the arm, detected erythema intensities and the calculated PASI scores can also be found here, as well as the used colour components which yielded the best lesion detection. The visual results of the other tests, i.e. surface meshes and point clouds are presented in Appendix A.

The disparity map in Figure 3b has a lot of noise in the background area, and so does the point cloud in Figure 3c. The detected lesion in Figure 3f seems to have holes in it. This can be seen again in the mesh in Figure 3g. The mesh in this figure also seems to have a lot of texture, different from the used stickers.

The results presented in Table I show that the method has some difference for every test. The smallest average difference is +4.18%, while the largest average difference is +141.86%. The average of all average differences is +37.45%. Notable is that the test with smallest difference of -0.47% (Figure 3), has many wrongly detected excess areas. Furthermore, the mesh of which the shape and detected excess areas closely resemble that of the picture taken, i.e. the mesh in Figure 7b, seems to have the biggest difference among the best scoring tests (+31.87\%).



(b) Mesh.

Fig. 4: Results of test with multiple red lesions on light skin tone, stereo pair 2.



Fig. 5: Results of test with red lesion on dark skin tone, stereo pair 3.

In Figure 4, the results of the second test of the multiple red lesions on the light skin can be seen. It can be seen that the lesions have been detected, but the mesh is not as rectangular

Lesion colour	Skin tone	Stereo pair	Used colour for detection	Calculated area (cm ²)	Actual area (cm ²)	Difference (%)	Average difference (%)	Area arm point cloud (cm ²)	Detected erythema intensity	PASI score
Red	Light	1	Cr	12.65	12	+ 5.43	+ 29.70	411.40	4	0
		2	Cr	11.94		- 0.47		483.80	4	0
		3	Cr	18.50		+ 54.13		411.74	4	0.8
		4	Cr	19.27		+ 60.59		479.12	4	0.8
		1	Cr	18.22	18	+ 1.23	+ 16.59	352.12	4	0.8
		2	Cr	18.19		+ 1.05		430.52	4	0.8
		3	Cr	18.39		+ 2.19		393.34	4	0.8
		4	Cr	29.14		+ 61.89		338.40	4	0.8
	Dark	1	Cr	12.79	12	+ 6.58	+ 4.18	488.91	4	0
		2	Cr	12.91		+ 7.65		381.59	4	0
		3	Cr	11.48		- 4.29		364.57	4	0
		4	Cr	12.81		+ 6.78		449.22	4	0
Pink	Light	1	Cb	11.03	12	- 8.06	5.06	453.25	2	0
		2	Cb	3.67		- 69.36		389.48	2	0
		3	Cb	13.85		+ 15.47		370.25	2	0
		4	Cb	17.00		+ 41.72		280.84	2	0.4
	Dark	1	Y	43.30	12	+ 217.03	+ 141.86	507.43	4	0.8
		2	Y	45.28		+ 277.30		585.17	4	0.8
		3	Y	16.95		+ 41.25		468.46	4	0
		4	Y	15.82		+ 31.87		410.87	4	0

TABLE I: Results of the calculated areas, detected erythema intensity and calculated PASI score.

as the stickers actually are. In Figure 5, the results of the third test of the red lesion on dark skin tone can be seen. The shape of this mesh resembles that of the used sticker. However, there are a few holes, which have been detected as excess area.



Fig. 6: Results of test with pink lesion on light skin tone, stereo pair 1.



(b) Mesh.

Fig. 7: Results of test with pink lesion on dark skin tone, stereo pair 4.

In Figure 6, the results of the first test of the pink lesion on light skin tone can be seen. The shape does not completely resemble that of the sticker, and areas in the middle of the mesh were detected as excess areas. In Figure 7, the results of the fourth test of the pink lesion on dark skin tone can be seen. There were no excess areas detected in the middle of the mesh, only at the outside. The shape resembles that of the sticker.

The MATLAB scripts used to obtain the results can be found in Appendix B.

V. DISCUSSION

As observed in the previous section, the calculated areas differ from the actual area. In general, it seems that the calculated area of the meshes is too large. This is because the obtained point clouds are too textured, resulting in a mesh with many peaks. Since the used stickers do not have any texture, it can be said that the point cloud and thus the mesh are inaccurate. This texture could perhaps be caused by quality of the disparity maps. These maps are not ideal and still have some pixels of different values in the area of interest. This would cause some points to be at a different depth than the others, explaining the peaks as seen in the mesh figures. Generally, disparity maps look worse for non-textured areas, which could also explain the noise in the background area. This noise is also present since the disparity range is not well defined for that area.

Detecting the lesion correctly does not always work. This could be caused by, for example, a wrongly taken picture, meaning part of the lesion is not visible in the pictures. Another possibility is that the sticker reflects light, which in the picture is seen as a lighter colour, making it more difficult to detect. This also causes holes in the point clouds. These holes could also be caused by occlusions in the disparity maps, which after filtering the point cloud results in even bigger occlusions, thus bigger holes in the lesion. Skin could also reflect light, making it particularly hard for light lesions to be detected, as the reflecting skin could be potentially seen as a lesion (e.g. Appendix A-E).

The detection of excess area does not work completely as intended. In many meshes the holes in the middle are coloured red when they should not be. Detecting these triangles should be further looked into. Another option is to look into other triangulation methods, which do not compute the convex hull, but just the correct area.

There cannot be said much about the accuracy of the calculated arm areas, but the results are likely to be inaccurate due to the point clouds being too textured. In every picture, the arm is not in the exact same position, which could explain why the values differ for every test. Using these areas to calculate the PASI scores by multiplying them by four defeats the whole purpose of why stereo vision is used, since doing so makes the end result imprecise. The calculated PASI scores have not been validated by comparing the results to existing methods. Furthermore, the predetermined areas are too small to calculate a PASI score, as for 12 cm² this would be equal to 0 anyway. It would be interesting to evaluate the method by looking at larger lesions. For the overall picture these scores have been added to the results table and could be used as reference for future work, to see how much the accuracy of the score improves for a more complete model.

As for the erythema intensity detection, comparing the reference images to the taken images, the results for the red lesions seem to be fair. For the pink lesion on light skin this detection also seems to work fine. However, for the pink lesion on the dark skin the detected erythema intensity is obviously incorrect. This is because for detecting this lesion the Y colours are examined for the point cloud as well as the reference images for the erythema. An image in Y colours is similar to a gray scale image, where the lightest colours are the brightest. Looking at the reference pictures in Figure 2, the skin of the person in Figure 2d has the lightest colour of all four images, and thus has the highest Y value. Comparing this to the lesion as bright as the one used for these tests, it is clear why the wrong intensity value was detected. Furthermore, it is not clear if this method will work for the other intensities, as this has not been tested.

From the tests, it seems that the method works the best for the red lesion on dark skin. However, only four pictures for each test have been processed, which is not enough to conclude on which colours the method works best.

VI. RECOMMENDATIONS

To improve the presented method, it should be investigated why the point clouds are textured more than they should be. Different disparity methods could be tested, and perhaps even different cameras. Cameras with a higher resolution could help with obtaining better and more detailed point clouds. Additionally, the method should be tested for more lesion colours, as in this paper only two colours have been tested. It would be even better if the method could be tested on actual lesions, not just stickers. Testing the method on the other body parts should be done as well. The erythema intensity detection could be improved by using deep learning, which would be more reliable than the current method presented in this paper. Furthermore, to make this method into an actual PASI determination method, the induration and desquamation intensity should be detected as well. For calculating the induration the point clouds have to be improved, as the depth of the lesion cannot be accurately determined if the point cloud has random peaks. For determining the scaling intensity, deep learning could be used here as well. Considering the area calculation, the method is not complete yet. Ideally, complete 360 degree point clouds of the human body parts are used. This could be realised by using multi-view stereo reconstruction. In addition, the point clouds should be examined for accuracy by comparing them to point clouds obtained from state-of-the-art camera setups.

VII. CONCLUSION

By using a stereo camera, a 3D model of a human arm can be made and its area can be calculated. Different coloured lesions present on the skin can be detected and examined for area. Using only a stereo camera and a computer makes the method portable and easy to use. Economically, this method would be significantly more affordable than existing reconstruction systems. However, many improvements should be done before the method can actually be used to accurately calculate PASI scores.

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APPENDIX A Additional results

In this appendix, the point clouds of all tests will be shown, together with their corresponding stereo pair.

A. Red lesion on light skin shade



(c) Stereo image 3.







Fig. 9: Point clouds of arm and lesion.



Fig. 10: Meshes of the detected lesions.

(c) Mesh 3.

B. Multiple red lesions on light skin shade





Fig. 11: Stereo images.

(c) Stereo image 3.



Fig. 12: Point clouds of arm and lesion.







Fig. 13: Meshes of the detected lesions.

C. Red lesion on dark skin shade



(a) Stereo image 1.

(b) Stereo image 2.



Fig. 14: Stereo images.

(c) Stereo image 3.



Fig. 15: Point clouds of arm and lesion.



(a) Mesh 1.







Fig. 16: Meshes of detected lesions.

D. Pink lesion on light skin shade



(a) Stereo image 1.

(b) Stereo image 2.



Fig. 17: Stereo images.

(c) Stereo image 3.

(d) Stereo image 4.









(a) Mesh 1.





Fig. 19: Meshes of the detected lesions.



E. Pink lesion on dark skin shade



(a) Stereo image 1.

(b) Stereo image 2.



Fig. 20: Stereo images.

(c) Stereo image 3.



Fig. 21: Point clouds of the arm and lesion.



(a) Mesh 1.







Fig. 22: Meshes of the detected point clouds.

APPENDIX B MATLAB SCRIPTS

In this appendix, the MATLAB scripts are presented. A description is added at the top of each function, clarifying what it does.

A. sr_functions.m

```
2 % This code detects lesions on body parts, using stereo images.
3 % The pictures may be taken beforehand
4
  % need to have the following functions in the path:
5
  % read_camera: take pictures and save them,
6
  % disparityRange: determine the disparity range of the stereo pair,
7
8
  % largestCluster: select the largest cluster of a point cloud,
   % processPtCloud: compute the denoised point cloud from stereo images,
9
  % surfArea3: compute the area of the mesh,
10
  % goodArea: compute the correct area of the mesh,
11
  % erythemaScore: determine the severity of the erythema (redness),
12
13
  % intensity: calculate intensity of the area,
14 % pasi: determine pasi score,
15
16
  % IMPORTANT NOTE: MAKE SURE THE STEREO CAMERA IS CALIBRATED AND THAT THE
17 % STEREOPARAMETERS are saved as 'stereoParams.mat' in the path
18
19
  clear all;
20 close all;
21 imtool close all;
22 set(gcf,'color','w');
23
  fontSize = 15;
24 tic % starts counting processing time
25
26
  q.newpic = questdlg('Would you like to take new pictures?',...
       'Yes'.'No'):
27
28
29
  % Handle response
30 switch q.newpic
31
      case 'Yes'
                    % take new pictures !! MAKE SURE OLD PICTURES ARE SAVED!!
          q.newp = 1;
32
      case 'No' % use old pictures; there should be pictures in the path
33
          q.newp = 0;
34
  end
35
36
  if q.newp == 1
37
  [LEFT,RIGHT,n] = read_camera;
38
39
  elseif q.newp == 0
40 n = 1:
41 % n is the number of stereo pairs to be processed, it's automatically
42 % determined if new pictures are taken
43 end
44
  %import the stereoparameters into the workspace
45
46
  load('stereoParams.mat'); % make sure the .mat file is in the right folder
47
  % read images from the workspace
48
49
  for i = 1 : n
50 stereoIm.LEFT.rgb{i} = imread(sprintf('TESTL_%s.png', num2str(i)));
st stereoIm.RIGHT.rgb{i} = imread(sprintf('TESTR_%s.png', num2str(i)));
  figure; imshow([stereoIm.LEFT.rqb{i},stereoIm.RIGHT.rqb{i}]);
52
53
  end
54
55 %% Determine disparity ranges
  disp("disparity");
56
  % disparity will be a cell containing all determined disparity ranges
57
58 disparity = disparityRange(n,stereoIm.LEFT.rgb,stereoIm.RIGHT.rgb,...
59
      stereoParams);
60 %% point cloud processing
61 disp("pointcloud");
  % point.all will be a cell containing all denoised point clouds
62
63 point.all = processPtCloud(n,stereoIm.LEFT.rgb,stereoIm.RIGHT.rgb,...
     stereoParams,disparity);
64
65
  %% delete invalid point clouds
   % makes figure full-screen
66
```

```
67 figure('units', 'normalized', 'outerposition', [0 0 1 1]);
68 for l = 1 : n
69 subplot (ceil (n/2), ceil (n/2), 1)
70 pcshow(point.all{l});
n t = sprintf('Point cloud %d', l);
72 title(t);
73
   end
74
   % if there's a faulty point cloud, this is the time to remove it
75
   prompt = sprintf(['Delete which point clouds? (type one number,' ...
76
77
        'space separated numbers, or leave the space empty)']);
   q.delnum = inputdlg(prompt);
78
  delete = str2num(q.delnum{1});
79
80
   selected = zeros(1,n);
                                   \% if n = 5, (0,0,0,0,0)
81
   for m = 1:n
82
       selected(m) = m;
                                   % (1,2,3,4,5)
83
   end
84
85
   % say we want to remove 2,3, selected = (1, 4, 5)
86
87 selected(delete) = [];
   for m = 1:size(selected,2)
89
   % pointSelect is only the 1st 4th and 5th point clouds
90
  pointSelect{m} = point.all{selected(m)};
91
   end
92
93
   for l = 1 : m
94
95
       subplot(ceil(m),ceil(m/2),l)
96
       pcshow(pointSelect{l});
       t = sprintf('Point cloud %d.', selected(l));
97
98
       title(t);
   end
99
100
   %% process point clouds
101
102
   for h = 1:size(pointSelect,2)
103
        % smooth point cloud arm
       pointArm = pointSelect{h};
104
       pointArmSmooth{h} = medfilt3(pointArm.Location,[5,1,7]);
105
106
        figure; pcshow(pointArmSmooth{h});
       tl = sprintf('Smoothed point cloud %d', h);
107
108
       title(tl);
109
       % Depending on the lesion colour, the point cloud colour will be chosen
110
        % to be further processed. Choose between Y Cb and Cr, look for which
111
        % colour the lesion is the brightest and contrasts the arm the most
112
113
       figure;
        % point cloud in Y
114
115
       colory = uint8(240*mat2gray(rgb2ycbcr(im2double(pointArm.Color))));
116
       pointArmYcbcr3{h} = pointCloud(pointArm.Location, 'Color',...
            [colory(:,1),colory(:,1),colory(:,1)]);
117
118
        subplot(2,2,1); pcshow(pointArmYcbcr3{h}); axis equal
       title('1');
119
120
121
       % point cloud in Cb
       colory = uint8(240*mat2gray(rgb2ycbcr(im2double(pointArm.Color))));
122
123
       pointArmYcbcr2{h} = pointCloud(pointArm.Location, 'Color', ...
            [colory(:,2),colory(:,2),colory(:,2)]);
124
125
        subplot(2,2,2); pcshow(pointArmYcbcr2{h}); axis equal
126
       title('2');
127
128
       % point cloud in Cr
129
       colory = uint8(240*mat2gray(rgb2ycbcr(im2double(pointArm.Color))));
       pointArmYcbcr1{h} = pointCloud(pointArm.Location, 'Color', ...
130
131
            [colory(:,3),colory(:,3),colory(:,3)]);
        subplot(2,2,[3,4]); pcshow(pointArmYcbcr1{h}); axis equal
132
133
       title('3');
134
       prompt = ['In which pointcloud is the lesion the brightest',...
135
            'and has the most contrast to the skin (1, 2 or 3)?'];
136
       a.l = inputdlg(prompt);
137
138
       q.light = str2num(q.l{1});
139
        if q.light == 1 %Y; works best for light lesions
140
            % look at YCBCR and construct lesion pointcloud
141
            pointArmYcbcr{h} = pointArmYcbcr3{h};
142
            range = 2;
143
```

```
144
            LES = pointArmYcbcr{h}.Color > max(max(colory(:,3))) - range;
        elseif q.light == 2 %CB
145
            % look at YCBCR and construct lesion pointcloud
146
147
            pointArmYcbcr{h} = pointArmYcbcr2{h};
            range = 25;
148
149
            LES = pointArmYcbcr{h}.Color > max(max(colory(:,2))) - range;
        elseif q.light == 3 %CR; works best for red lesions
150
            % look at YCBCR and construct lesion pointcloud
151
            pointArmYcbcr{h} = pointArmYcbcr1{h};
152
            range = 60;
                            % 30 for dark skin, 60 for lighter
153
154
            LES = pointArmYcbcr{h}.Color > max(max(colory(:,3))) - range;
155
        end
156
157
        [ROWS, COLS] = find(LES == 1); % obtain the rows with the right colour
       pointLesionRgb = select(pointArmYcbcr{h},ROWS); % make the point cloud
158
        figure; pcshow(pointLesionRgb); axis equal
159
       title('Pointcloud of lesion before filtering');
160
161
162
       % denoise pointcloud
163
        % outlier threshold. A point is considered an outlier if the
        % average distance to numNeighbors > threshold
164
       pointLesionDenoised{h} = pcdenoise(pointLesionRgb, 'NumNeighbors', ...
165
            100, 'Threshold', 0.01); % removes outliers from the pointcloud
166
167
        figure;pcshow(pointLesionDenoised{h}); axis equal;
       title('Denoised pointcloud lesion')
168
169
170
        % if there are clusters of which the lesion is the largest, this can be
        % filtered out
171
172
       q.cluster = questdlg('Use cluster detection?',...
173
                                 'Yes'.'No'):
        % Handle response
174
175
       switch q.cluster
            case 'Yes'
                                 % will detect largest cluster
176
177
               q.cl = 1;
            case 'No'
                                 % nothing changes
178
                q.cl = 0;
179
180
        end
181
       if q.cl == 1
182
183
            pointLesionCluster = largestCluster(0.02,pointLesionDenoised{h});
            figure;pcshow(pointLesionCluster);
184
185
            title('Cluster detected lesion');
       else
186
            pointLesionCluster = pointLesionDenoised{h};
187
        end
188
189
190
        % smooth lesion surface by using median filter
       pointLesionSmooth{h} = medfilt3(pointLesionCluster.Location, [3,1,3]);
191
192
        figure;pcshow(pointLesionSmooth{h},'r');
193
       title('lesionsmooth');
194
195
        % delaunay triangulation of whole pointcloud
        disp("triangulation arm");
196
107
       pointcloudTempA = pointArmSmooth{h};
        % obtain x y and z coordinates of the vertices
198
       triangulation.armX = double(pointcloudTempA(:,1));
199
200
       triangulation.armY = double(pointcloudTempA(:,2));
       triangulation.armZ = double(pointcloudTempA(:,3));
201
202
203
        % delaunay outputs a matrix of m x 3 where m is the number of triangles
       triangulation.arm = delaunay(triangulation.armX, triangulation.armY);
204
205
        figure; triplot(triangulation.arm,triangulation.armX,...
            triangulation.armY);
206
207
208
        % surface mesh
       figure; trisurf(triangulation.arm,triangulation.armX,...
209
210
            triangulation.armY,triangulation.armZ, colory(:,3)); axis equal;
        xlabel('x');
211
       ylabel('y');
212
213
        zlabel('z');
       caption = sprintf('Surface meshed point cloud');
214
215
       title(caption, 'FontSize', fontSize);
216
        % triangulation of lesion
217
       disp("triangulation lesion");
218
       pointcloudTempL = pointLesionSmooth{h};
219
```

% obtain x y and z coordinates of the vertices

220

```
triangulation.lesionX = double(pointcloudTempL(:,1));
221
        triangulation.lesionY = double(pointcloudTempL(:,2));
222
        triangulation.lesionZ = double(pointcloudTempL(:,3));
223
224
        % delaunay outputs a matrix of m x 3 where m is the number of triangles
225
226
        triangulation.lesion = delaunay(triangulation.lesionX,...
            triangulation.lesionY);
227
        figure; triplot(triangulation.lesion,triangulation.lesionX,...
228
            triangulation.lesionY);
229
230
231
        % surface mesh
        figure; trisurf(triangulation.lesion,triangulation.lesionX,...
232
            triangulation.lesionY,triangulation.lesionZ, ...
233
234
            pointLesionCluster.Color(:,3)); axis equal;
        set(gcf, 'Renderer', 'zbuffer');
235
        xlabel('x');
236
        ylabel('y');
237
        zlabel('z');
238
239
        caption = sprintf('Surface meshed point cloud');
        title(caption, 'FontSize', fontSize);
240
241
242
        % calculate surface area
243
244
       disp("area arm");
        % obtain the area, the colored triangles and their vertice coordinates
245
        [surface.arm] = surfArea3(triangulation.arm, triangulation.armX,...
246
247
           triangulation.armY, triangulation.armZ);
        area.arm{h} = sum(surface.arm); % total area, including wrong triangles
248
249
        axis equal
250
        %%% NOTE: AREA IS IN SQUARE DECIMETERS %%%
251
252
        % remove the excess triangles
253
254
        disp("correctArea arm");
255
        figure; trisurf (triangulation.arm, triangulation.armX, ...
256
257
            triangulation.armY,triangulation.armZ,...
            pointArmYcbcr{h}.Color(:,3)); hold on;
258
259
        % goodArea selects which triangles do not belong
260
        surface.armCorrect = goodArea(triangulation.arm, triangulation.armX,...
            triangulation.armY, triangulation.armZ, surface.arm, ...
261
262
            pointArmYcbcr{h},17,2);
263
        % total area excluding the large excess triangles
264
265
        area.armCorrectDM{h} = sum(surface.armCorrect);
       area.armCorrectCM{h} = area.armCorrectDM{h} * 100; % in cm<sup>2</sup>
266
267
        % area lesion
268
269
       disp("area lesion");
270
        % obtain the area, the colored triangles and their vertice coordinates
        [surface.lesion] = surfArea3(triangulation.lesion, ...
271
272
            triangulation.lesionX, triangulation.lesionY, ...
            triangulation.lesionZ);
273
274
        area.lesion{h} = sum(surface.lesion); % total area of triangulation
275
        axis equal
276
277
        disp("correctArea lesion");
        figure; trisurf (triangulation.lesion, triangulation.lesionX, ...
278
279
            triangulation.lesionY,triangulation.lesionZ,...
280
            pointLesionCluster.Color(:,3)); hold on;
        title(sprintf('Meshed lesion %d',h));
281
282
        surface.lesionCorrect = goodArea(triangulation.lesion,...
            triangulation.lesionX, triangulation.lesionY, ...
283
            triangulation.lesionZ, surface.lesion, ...
284
285
            pointLesionCluster,3,5); axis equal;
        % for multiple lesions, change 3,5 to 4,3
286
287
        % compute total area excluding the large excess triangles
288
        area.lesionCorrectDM{h} = sum(surface.lesionCorrect);
289
        area.lesionCorrectCM{h} = area.lesionCorrectDM{h} * 100; % in cm
290
291
  end
292
293
   %% process areas
294 % plot the point clouds of the arm and the lesion in one figure
  figure('units', 'normalized', 'outerposition', [0 0 1 1]);
295
   for i = 1:h
296
297
        if h == 1 % for one stereo pair, plot the clouds next to eachother
```

```
298
            subplot(1,2,1)
           pcshow(pointArmYcbcr{i});
299
            title('Point cloud of arm in Cr.')
300
            subplot(1,2,2);
301
            pcshow(pointLesionSmooth{i},'w');
302
303
            title('Point cloud of detected lesion.')
       else
304
            subplot(ceil(h/2),ceil(h),i)
305
            pcshow(pointArmYcbcr{i});
306
            subplot(ceil(h/2),ceil(h), h + i)
307
308
            pcshow(pointLesionSmooth{i}, 'w');
       end
309
  end
310
311
   % calcuate the average of the arm and lesion areas
312
313
   sumA = 0;
   sumL = 0;
314
   for p = 1:h
315
316
       sumA = sumA + area.armCorrectDM{h};
317
       sumL = sumL + area.lesionCorrectDM{h};
318
  end
  area.armAverage = sumA/h;
319
320 area.lesionAverage = sumL/h;
321
   % assume that the area of the arm as calculated here is about half of the
322 % total. So for two arms, area_good*4
323
324
325 % how much area the wound is compared to the arm (in percent)
326 %F = (area.lesionAverage/area.armTotal)*100;
327
328 for i = 1:n
329 % return intensity score (0-4)
330 area.armTotal = area.armCorrectCM{i}*4;
331 F = (area.lesionCorrectCM{i}/area.armTotal) * 100;
332 arm.intensity.area{i} = intensity(F);
333
334
   % determine erythema severity
  arm.intensity.erythema{i} = erythemaScore(pointLesionDenoised{i},q.light);
335
  %pointLesionDenoised is the YCBCR colored cloud of the lesion
336
337
338 % calculate pasi score
339 PASI.head = [0,0,0,0];
                             % (area, erythema, induration, desquamation)
340 PASI.arms = [arm.intensity.area{i},arm.intensity.erythema{i},0,0];
341 PASI.trunk = [0,0,0,0];
342 PASI.legs = [0,0,0,0];
343 score{i} = pasi(PASI.head,PASI.arms,PASI.trunk,PASI.legs);
344
345 % show pasi score
346 PASIscore = sprintf('PASI score %d: %d', i, score{i});
347
  box.pasi = msgbox({PASIscore}, 'PASI score');
  disp(sprintf('Lesion area = %d', area.lesionAverage))
348
349
  end
350
   % calculate difference between areas
351
  prompt = 'What is the actual area (in square cm)?';
352
   q.area = inputdlg(prompt);
353
354
   for i = 1:n
       difference{i} = str2num(q.area{1})-area.lesionCorrectCM{i};
355
       diffp = round(((abs(difference{i}))/str2num(q.area{1})) * 100,2);
356
357
       if difference{i} < 0 % the calculated area is bigger</pre>
           text.diff = sprintf(['The calculated area %d is %f percent',...
358
359
                'larger than the actual area'], i, diffp);
       else % calculated area is smaller
360
            text.diff = sprintf(['The calculated area %d is %f percent',...
361
362
                'smaller than the actual area'], i, diffp);
       end
363
       text.qarea = sprintf('The actual area is %f square cm',...
364
           str2num(q.area{1}));
365
       text.carea = sprintf('The calculated area is %f square cm',...
366
           area.lesionCorrectCM{i});
367
       box.area = msgbox({text.garea, text.carea, text.diff});
368
  end
369
370
   totalTime = toc
```

```
i function [LEFT,RIGHT,n] = read_camera
2 % read_camera determines the area of a meshed point cloud
   % area = goodArea(triangulation, X, Y, Z, A, pointcloud, length, factor))
3
4 % - area is the area of the meshed point cloud
  % - triangulation is the triangulated point cloud
   % - X, Y, Z are the coordinates of the vertices of the triangles
6
   \ensuremath{\$} - A is the area of all triangles summed up
7
   % - pointcloud is the pointcloud of which the triangulation is made
8
   \ensuremath{\$} – length is an integer; the maximum line length is Xlim/length
9
  \, - factor is an integer, indicating how much larger a triangle may be
10
11 % wrt the mean area
12 % !!! BEFORE RUNNING THIS CODE MAKE SURE EXISTING IMAGES WON'T BE
13
   % OVERWRITTEN!!!!
14 clear all
15 close all
16
17 camList = webcamlist;
                                   % shows the list of available cameras
18 \text{ cam} = \text{webcam}(2);
                                    % make sure to select the stereo camera!
19 preview(cam);
                                    % gives a real-time view of the camera
20 fontSize = 10;
21
22 n = 0;
   figure;
23
  while 1
24
       %ask user to take picture
25
26
       user = input('Take picture? yes = y, no = n ', 's');
       if user == 'n'
27
           if n == 0
28
               f = errordlg('No pictures were taken.');
29
30
           %if the user wants to stop taking pictures
           end
31
32
           break
       else
33
           %user wants to take a picture
34
          n = n + 1;
35
          picture = snapshot(cam); % takes a picture
36
37
           temp = picture;
           imshow(temp);
                                        % view the pictures before saving them
38
39
           % save pictures
           LEFT{n} = picture(:,1:(size(picture,2))/2,:);
40
           RIGHT{n} = picture(:,((size(picture,2))/2)+1 : size(picture,2),:);
41
       end
42
43
       X = ['You have taken ', num2str(n) ,' picture(s).'];
       disp(X);
44
45
  end
46
47 %show and save the pictures as .png
48
49
   figure;
   for k = 1:n
50
       if n \leq 2
                                % i.e. for only 1 or two pictures
51
          subplot(n, 1, k);
52
       else
53
         subplot(ceil(n/2),ceil(n/2),k);
54
55
       end
56
       %write the pictures to .png files
       imshow([LEFT{k}, RIGHT{k}]);
57
       imwrite(LEFT{k},sprintf('TESTL_%d.png',k));
58
       imwrite(RIGHT{k}, sprintf('TESTR_%d.png',k));
59
       caption = sprintf('Stereo pair #%d of #%d', k, n);
60
61
       title(caption, 'FontSize', fontSize);
       hold on
62
63
   end
64
65
  end
```

```
i function disparity = disparityRange(n,LEFT,RIGHT,stereoParams)
2 % disparityRange determines the disparity range for stereo pairs, if the
   % user wants to make a disparity map of a subject in the foreground.
3
4 % disparity = disparityRange(n,LeftIm,RightIm,stereoParameters)
  % - disparity a cell containing the disparity ranges for each stereo pair
5
6~ % - n is the amount of stereo pairs used as input
  \ensuremath{\$} - LEFT is a cell containing all left images
7
  % - RIGHT is a cell containing all right images
8
  \ensuremath{\$} - stereoParameters is the parameters obtained from the stereo
9
10
  % calibration app
11
12 for pair = 1: n
        % rectify the images
13
       [IL{pair}, IR{pair}] = rectifyStereoImages(LEFT{pair}, RIGHT{pair}, ...
14
           stereoParams);
15
       I = stereoAnaglyph(IL{pair},IR{pair});
16
      figure; imshow(I);
17
18
       msgbox(sprintf(['Please draw a line from a lesion point in the'...
           'left image to the same point in the right image. Then,'...
19
           'double click the line.']));
20
      % draw a line on the image
21
       h = imline;
22
       position = wait(h); % then double click on the line
23
24
25
       \ obtain 2x2 matrix containing x and y coordinates of the endpoints
26
       pos = getPosition(h);
       XL = pos(1, 1);
27
       YL = pos(1, 2);
28
       XR = pos(2, 1);
29
30
       YR = pos(2, 2);
31
       % calculate the lenght of this line
32
       length = ceil(sqrt((XR-XL)^2+(YR-YL)^2));
33
       DISPARITY{pair} = [length - 20, length + 68];
34
35 end
36 disparity = DISPARITY;
```

```
i function ptCloud = largestCluster(minDistance, pointcloud)
2 % largestCluster filters the input pointcloud and returns the largest
3 % cluster
4 % ptCloud = largestCluster(minDistance, pointcloud)
5 % - ptCloud = output point cloud
6 % - minDistance is the minimum Euclidean distance between points from
\tau % two different clusters, specified as a positive scalar
8 % - pointcloud is the input point cloud
10 % Segment the point cloud; detect the clusters
11 [labels,numClusters] = pcsegdist(pointcloud,minDistance);
12 figure;
pcshow(pointcloud.Location, labels)
14 colormap(hsv(numClusters))
15 title('Point Cloud Clusters')
16
\scriptstyle 17 % remove unwanted clusters, keep the largest
18
  % pointcloud.Location contains as much points as there are labels
  % connect the indices of the points from labels containing number M, to
19
20
  % the indices of input pointcloud
                           % finds the most occuring number in labels;
21 M = mode(labels);
                           % The Mth cluster is the biggest cluster
22
23 F = find(labels == M); % returns an array of indices containing M
24 ptCloud = select(pointcloud,F); % select only those points
25 end
```

```
1 function pointcloud = processPtCloud(n,LEFT,RIGHT,stereoParams,dispRange)
2 % processPtCloud returns the point cloud of a subject in the foreground.
   % pointcloud = processPtCloud(n,LEFT,RIGHT,stereoParams,dispRange)
3
  % - pointcloud is the denoised pointcloud
4
5 % - n is the amount of stereo pairs used as input
  % - LEFT is a cell containing all left images
6
  % - RIGHT is a cell containing all right images
7
  % - stereoParameters is the parameters obtained from the stereo
  % calibration app
9
10 % - dispRange is the disparity range (e.g. [0,64]), should be a cell
11
12 fontSize = 15;
13
  for m = 1 : n
14
15
       % rectify the images
       [IL{m},IR{m}] = rectifyStereoImages(LEFT{m},RIGHT{m},stereoParams);
16
17
       figure;
       subplot(1,2,1);
18
       IL{m} = histeq(IL{m});
                                      % preprocess images
19
       ILB{m} = rgb2gray(IL{m});
20
      imshow(ILB{m});
21
22
23
       subplot(1,2,2);
       IR\{m\} = histeq(IR\{m\});
                                      % preprocess images
24
       IRB{m} = rgb2gray(IR{m});
25
26
       imshow(IRB{m});
27
28
       A = stereoAnaglyph(IL{m},IR{m}); % makes a red-cyan overlapping image
       figure; imshow(A);
29
       caption = sprintf('Anaglyph picture %d', m);
30
       title(caption, 'Fontsize', fontSize);
31
32
33
       disparityRange = dispRange{m};
       uniqueness = 11; % low value --> less reliable disparity map
34
35
36
       disparityMap1{m} = disparitySGM(ILB{m}, IRB{m}, ...
           'DisparityRange', disparityRange, 'UniquenessThreshold',...
37
38
           uniqueness);
39
       % remove noise in the disparity map
40
       disparityMap{m} = medfilt2(disparityMap1{m});
41
       figure; imshow(disparityMap{m},disparityRange);
42
43
       caption = sprintf('Disparity Map of picture %d', m);
       title(caption, 'Fontsize', fontSize);
44
45
       colormap(gca,jet);
46
       colorbar;
47
48
       %obtain the 3D points from the disparity maps
       points3D = reconstructScene(disparityMap{m}, stereoParams);
49
50
       pointCl{m} = removeInvalidPoints(pointCloud(points3D ./ 100,...
           'Color', IL{m})); % remove NaN and Inf points
51
       figure; pcshow(pointCl{m});
52
53
       caption = sprintf('Point cloud #%d of #%d', m, n);
       title(caption, 'FontSize', fontSize);
54
55
56
       % preprocess the point cloud
       % downsampling
57
       ptClDown{m} = pcdownsample(pointCl{m}, 'gridAverage', 0.01);
58
       figure; pcshow(ptClDown{m});
59
       caption = sprintf(['Downsampled point cloud #%d of #%d,'...
60
61
           'gridsize 0.01'], m, n);
       title(caption, 'FontSize', fontSize);
62
63
       % preprocess the point cloud
64
65
       % denoising
       % filter by looking at the distance between point clouds
66
       numNeighbors = 100; % number of nearest neighbor points
67
       % outlier threshold. A point is considered an outlier if
68
69
       % the avg distance to numNeighbors > threshold
70
       threshold = 0.01;
       ptCldn{m} = pcdenoise(ptClDown{m}, 'NumNeighbors', numNeighbors,...
71
           'Threshold', threshold); % removes outliers from the pointcloud
72
73
       figure;
```

```
pcshow(ptCldn{m});
74
75
        % denoising
76
        % Segment the point cloud; detect the clusters
minDistance = 0.015;  % should be bigger than 'gridAverage'
77
78
        point{m} = largestCluster(minDistance,ptCldn{m}) ;
79
80
        figure; pcshow(point{m});
81
        caption = sprintf('Downsampled denoised point cloud #%d of #%d', m, n);
title(caption, 'FontSize', fontSize);
82
83
84
85
86 end
87
88 % write point cloud to .mat
89 pointcloud = point;
90 end
```

```
1 function [area] = surfArea3(triangulation, X, Y, Z)
_{\rm 2} % surfArea3 returns the area of a triangulation; all triangles summed
3 % area = surfArea3(triangulation, X, Y, Z)
4 % area = the area of the input triangulation
5 % - triangulation is the triangulated surface,
6 % i.e. triangulation = delaunay(X,Y)
7 \, \, - X,Y,Z are the x y and z coordinates of the vertices
8
9 tri = triangulation;
10 sizetri = size(tri,1);
11
12 % total area
13 A = [];
14
15 for i = 1:sizetri
                      % triangle #
      tri_num = i;
16
       % each row of tri contains the #th point from the point cloud of which
17
18
      % a triangle is made
       % get their coordinates
19
      tx = X(tri(tri_num,:)); % tx is the x coordinates of the three points
20
      ty = Y(tri(tri_num,:));
21
      tz = Z(tri(tri_num,:));
22
23
      point1 = [tx(1,:),ty(1,:),tz(1,:)]; %xyz coordinates of point 1
24
25
      point2 = [tx(2,:), ty(2,:), tz(2,:)];
      point3 = [tx(3,:),ty(3,:),tz(3,:)];
26
27
      % surface area
28
      %fill3(x,y,z,'r')
x = tx(:)';
29
30
                                    % transpose tx, ty, tz. x = 1x3 matrix
       y = ty(:)';
31
       z = tz(:)';
32
       ons = [1 1 1];
33
       A(i) = 0.5*sqrt(det([x;y;ons])^2 + det([y;z;ons])^2 + ...
34
35
           det([z;x;ons])^2);
36 end
37
  area = A;
38
39 end
```

```
1 function area = goodArea(triangulation, X, Y, Z, A, pointcloud, length, factor)
2 % goodArea determines the area of a meshed point cloud
   % area = goodArea(triangulation, X, Y, Z, A, pointcloud, length, factor))
3
4 % - area is the area of the meshed point cloud
  % - triangulation is the triangulated point cloud
   % - X, Y, Z are the coordinates of the vertices of the triangles
6
   \ensuremath{\$} - A is the area of all triangles summed up
7
   % - pointcloud is the pointcloud of which the triangulation is made
8
  \ensuremath{\$} - length is an integer; the maximum line length is Xlim/length
9
  % - factor is an integer, indicating how much larger a triangle may be
10
11 % wrt the mean area
12
13
14 tri = triangulation;
15 sizetri = size(tri,1);
16 meanA = mean(A);
17 A_good = [];
18 W = []; % the Wth triangle is wrong
19 xlim = abs(pointcloud.XLimits(2)-pointcloud.XLimits(1))/length;
20
21
   for i = 1: sizetri
      tri_num = i; % triangle #
22
       % look at the length of the triangle edges
23
       % if one edge is longer than half of the lesion, it is a wrong triangle
24
25
       % tx is the x coordinates of the three points (3x1 matrix)
26
      tx = X(tri(tri_num,:));
27
       ty = Y(tri(tri_num,:));
28
      tz = Z(tri(tri_num,:));
29
30
       point1 = [tx(1,:),ty(1,:),tz(1,:)]; %xyz coordinates of point 1
31
32
       point2 = [tx(2,:), ty(2,:), tz(2,:)];
       point3 = [tx(3,:),ty(3,:),tz(3,:)];
33
34
       % calculate length of triangle edges
35
       d = ((x^2 - x^1)^2 + (y^2 - y^1)^2 + (z^2 - z^1)^2)^{1/2}
36
       length12 = (sqrt((point2(1)-point1(1))^2+(point2(2)-point1(2))...
37
           + (point2(3)-point1(3))^2));
38
       length23 = (sqrt((point3(1)-point2(1))^2+(point3(2)-point2(2))...
39
           + (point3(3)-point2(3))^2));
40
       length13 = (sqrt((point3(1)-point1(1))^2+(point3(2)-point1(2))...
41
           +(point3(3)-point1(3))^2));
42
43
       if A(i) > meanA*factor % means the triangle does not belong
44
45
          if length12 > xlim || length23 > xlim || length12 > xlim
               W(end+1) = i;
46
47
          end
       else
48
           A_good(end + 1) = A(i);
49
50
       end
51
52 end
53 area = A_good;
54
  % plot wrong triangles
55
   % get their coordinates
56
57 tx = X(tri(W,:));
58 ty = Y(tri(W,:));
59 tz = Z(tri(W,:));
   % plot specific triangles
60
61 \text{ txc} = [tx, tx(:, 1)];
                               %close them for plotting
62 tyc = [ty, ty(:,1)];
   tzc = [tz, tz(:, 1)];
63
64 fill3(txc.', tyc.', tzc.', 'r');
65 hold off
66
   end
```

```
1 function intensity = erythemaScore(pointcloud, colour)
2 % erythemaScore determines the erythema intensity of the lesion point cloud
   % intensity = erythemaScore(pointcloud, colour)
3
4 % - intensity is an integer between 1 and 4
s % - pointcloud is the input point cloud of the lesion, in Y Cb or Cr
_{\rm 6} % - colour is one of the YCbCr components, same as the lesion point cloud
8 % look at whether the values of the point cloud colors are in one of the
  % four intensities
9
10
  figure;
11
  for i = 1 : 4
12
       % read reference images
13
       EryRGB{i} = imread(sprintf('erythema_%s.PNG', num2str(i)));
14
       % transform them into ycbcr colour space
15
       YCBCR = rgb2ycbcr(EryRGB{i});
16
       COLOUR{i} = YCBCR(:,:,colour); % either y cb or cr
17
18
       % look at the maximum values
       MAX = max(max(COLOUR{i}));
19
20
       C = double(COLOUR{i} > MAX-60);
       CMAX{i} = uint8(C .* double(COLOUR{i}));
21
       subplot(2,2,i); imshow(CMAX{i});
22
       CM = CMAX{i};
23
       range{i} = [min(CM(CM > 0)), max(CM(CM > 0))];
24
25 end
26
27 erythema1 = [];
28 erythema2 = [];
29 erythema3 = [];
30 erythema4 = [];
31
32 range1 = range{1};
33 range2 = range{2};
_{34} range3 = range{3};
35 range4 = range{4};
36
   for k = 1:size(pointcloud.Color,1) % for every point in the pointcloud
37
       RED = pointcloud.Color(k,3);
38
39
       % if the colour of the point falls into the range
40
       if RED \geq range1(1) && RED \leq range1(2)
41
           erythemal(end+1) = 1;
42
       end
43
       if RED \geq range2(1) && RED \leq range2(2)
44
45
           erythema2(end+1) = 1;
       end
46
       if RED \geq range3(1) && RED \leq range3(2)
47
           erythema3(end+1) = 1;
48
       end
49
       if RED \geq range4(1) && RED \leq range4(2)
50
           erythema4(end+1) = 1;
51
       end
52
53
54 end
_{55} % determine which index has the highest value
  summ = [sum(erythema1), sum(erythema2), sum(erythema3), sum(erythema4)];
56
   [M,I] = max(summ);
57
58 intensity = I;
59 end
```

```
I. intensity.m
```

```
i function IArea = intensity(F)
_{2}\, % intensity determines the intensity score of the area in percentages
3 % IArea = intensity(F)
4 % IArea = intensity score of the area (1-4)
s\, % F = input percentage of lesion area to body area
6 if F < 90
       if F < 70
7
           if F < 50
8
                if F < 30
9
                     if F < 10
10
                         if F < 1
11
                              IArea = 0;
12
13
                         else
                              IArea = 1;
14
15
                         end
                     else
16
                         IArea = 2;
17
                     end
18
                else
19
                     IArea = 3;
20
                end
21
           else
22
23
                IArea = 4;
           end
24
       else
25
            IArea = 5;
26
       end
27
28
  else
       IArea = 6;
29
30
31 end
32 end
```

```
1 function score = pasi(head,arms,trunk,legs)
2 % pasi calculates the PASI score from input intensity scores
3 % score = pasi(head,arms,trunk,legs)
4 \, \, - score is the calculated PASI score
s\, % - head is an array containing the intensity scores of the four criteria
6 % head = [area,erythema,induration,desquamation]
\tau % – arms is the same as head, but then for the arms
  % - trunk is the same as head, but then for the trunk
8
  9
10
11 % B1 = A1 * 0.1;
12 % B2 = A2 * 0.2;
  % B3 = A3 * 0.3;
13
  % B4 = A4 * 0.4;
14
  ŝ
15
  % C1 = B1 * Area;
16
  % C2 = B2 * Area;
17
18
  % C3 = B3 * Area;
  % C4 = B4 * Area;
19
20
  2
21 \% \text{ score} = C1 + C2 + C3 + C4;
22
23
  %head = [area,erythema,induration,desquamation]
24
25 A1 = sum(head(:,2:4));
                             % sum all intensities except area
26 A2 = sum(arms(:,2:4));
27 A3 = sum(trunk(:,2:4));
28 A4 = sum(legs(:,2:4));
29
  B1 = A1 * 0.1;
30
B2 = A2 * 0.2;
32 B3 = A3 * 0.3;
  B4 = A4 * 0.4;
33
34
35
  C1 = B1 * head(1);
36 C2 = B2 \star arms(1);
37 C3 = B3 * trunk(1);
38 C4 = B4 * legs(1);
39
40 score = C1 + C2 + C3 + C4;
41 end
```